Is the number of antral follicles an interesting selection criterium for fertility in cattle?

F. Morotti¹, T.R.R. Barreiros², F.Z. Machado¹, S.M. González¹, L.S.R. Marinho¹, M.M. Seneda^{1,3}

¹Laboratório de Reprodução Animal, DCV-CCA-UEL, Londrina, PR, Brazil. ²Laboratório de Biotecnologia da Reprodução Animal, DVPA, UENP, Bandeirantes, PR, Brazil.

Abstract

Recent studies indicate that the antral follicle population may be of paramount importance to improve reproductive performance in cows. There is already an agreement that the antral follicle count (AFC; follicles \geq 3 mm in diameter) is a highly variable trait among animals, but with high repeatability in the same individual. Thus, females can be classified into low, intermediate or high AFC. Several studies in Bos taurus show a positive correlation between AFC and fertility parameters, such as increased quantity and quality of embryos, better pregnancy rates, higher progesterone levels, among others. However, there is still no consensus on AFC in Bos indicus females and indicustaurus. This article aims to discuss the main aspects related to the population of antral follicles and its relation to the reproductive performance associated with the most common techniques in assisted reproduction (timed artificial insemination, in vitro embryo production, embryo transfer and superovulation).

Keywords: antral follicle count, embryo production, follicular dynamics, pregnancy rate, ultrasonography.

Introduction

High genetic quality animals can be multiplied efficiently using reproductive biotechnologies such as artificial insemination and embryo production. These biotechnologies are useful strategies and known worldwide for improving genetics and productivity of flocks over a short period (Mapletoft and Hasler, 2005; Boni, 2012; Hansen, 2014).

Recently, there have been an increasing interest in studies concerning antral follicle count (AFC) and its influence on the reproductive performance in cattle, as well as its applications in reproductive biotechnologies (Ireland *et al.*, 2011; Pontes *et al.*, 2011; Rico *et al.*, 2012; Silva-Santos *et al.*, 2014a, b). Such fact may result in immense repercussions on the current scenario of animal reproduction, considering the significant increase in the world's embryo production. Despite several favorable results about AFC in *Bos taurus*, many aspects of reproductive physiology remain unknown. Considering AFC in *Bos indicus* there are many points to be addressed, particularly the impact on fertility when using *in vitro* embryo production (IVEP), timed artificial insemination (TAI) and timed embryo transfer (TET).

The high variability in the population of antral follicles is a hallmark in cattle (Burns *et al.*, 2005) with low, intermediate or high AFC (Santos *et al.*, 2012, 2013; Mendonça *et al.*, 2013). Despite the high variability among animals, there is a high repeatability of the number of follicles observed in the same individual through evaluations carried out during a period (Burns *et al.*, 2005; Ireland *et al.*, 2007, 2008, 2009).

This constancy in AFC in the same individual becomes a strategic resource for the possibility of classifying an animal by the AFC with a single ultrasound examination. For taurus animals, AFC is directly correlated with the size of the ovarian follicular reserve (Ireland et al., 2011), which was not proven in indicus females, considering fetuses, heifers and cows (Silva-Santos et al., 2011). However, other factors such as genetics (Walsh et al., 2014), maternal environment, nutritional status and healthiness (Ireland et al., 2011; Evans et al., 2012) also appear to influence the AFC. For example, the nutritional status and the metabolic rate were mentioned as factors which affect the follicular growth, oocyte quality and secretion of reproductive hormones in cattle (Jimenez-Krassel et al., 2009; Mossa et al., 2010; Evans et al., 2012).

The AFC may also influence the production of cattle embryos, both *in vivo* and *in vitro*, especially as the number of embryos produced by donor but also in process efficiency, with higher rates for high AFC animals (Ireland *et al.*, 2008; Santos *et al.*, 2014; Silva-Santos *et al.*, 2014a).

The importance of the AFC and its relationship with pregnancy rates must also be emphasized. Studies conducted with *taurus* females had higher pregnancy rates for high AFC females (Cushman *et al.*, 2009; Evans *et al.*, 2012; Mossa *et al.*, 2012). However, in recent studies with *indicus-taurus* and *indicus* animals, a better performance regarding pregnancy rates was not observed in high-AFC animals (Mendonça *et al.*, 2013; Santos *et al.*, 2014). Surprisingly, some data suggest a better performance regarding pregnancy rates for low AFC cows (Santos *et al.*, 2013).

In addition to ultrasound, the measurement of the concentration of anti-Mullerian Hormone (AMH)

³Corresponding author: mseneda@uel.br Phone: +55(43)3371-5622; Fax: +55(43)3371-4063 Received: May 27, 2015 Accepted: July 13, 2015

can also be used for selection of females according to the size of follicular population, since AMH has been considered as a factor highly correlated with the number of antral follicles and healthy oocytes (Hehenkamp *et al.*, 2006; Ireland *et al.*, 2011).

Considering the impact of reproductive biotechnology in genetic improvement and doubts as to the effect of antral follicle count on the reproductive performance in *indicus* animals, this review aims to discuss the main aspects of the AFC related to the number of antral follicles and its relationship with follicular dynamics, TAI, OPU/IVF and SOV.

Antral follicles count and its repeatability

The number of ovarian antral follicles is highly variable in different species (Burns *et al.*, 2005; Ireland *et al.*, 2007; Pontes *et al.*, 2011; Santos *et al.*, 2012), but in cattle, there is a high repeatability in the same individual (Burns *et al.*, 2005). Thus, the possibility of selecting *taurus* females by ultrasonography based on the AFC was demonstrated, due to the high degree of repeatability between individuals, regardless of race, age, breeding season, lactation or pregnancy conditions (Burns *et al.*, 2005). The same situation of AFC repeatability was demonstrated in *indicus* blooded animals; evaluating the same females for several months it was possible to identify the high correlation (0.90-0.92) in *taurus-indicus* weaned and yearling females (Santos *et al.*, 2012; Silva-Santos *et al.*, 2014a).

Therefore, a single routine ultrasound examination can identify females with low, intermediate or high AFC, according to findings also observed in Nelore females (Morotti *et al.*, 2015; DCV-CCA-UEL, Londrina, PR, Brazil; unpublished data).

Antral follicles count and embryo production

Considering the importance of the production of embryos for genetic improvement of livestock, it is noted that there is a great variability in the number of embryos produced by the donor, in both in vitro and in vivo methods (Pontes et al., 2009, 2010, 2011). The high variability in oocyte recovery rate and superovulatory response are important factors that affect the success of the bovine embryo production (Taneja et al., 2000; Ireland et al., 2007, 2011; Silva-Santos et al., 2014a). In this context, several studies with taurus animals and some studies with indicus-blooded animals have shown that a greater number of follicles is associated with quantitative benefits to the success of IVEP and SOV (Taneja et al., 2000; Singh et al., 2004; Ireland et al., 2008; Silva-Santos et al., 2014a). For *indicus-taurus* animals, the average number of embryos per collection was also higher for females with high compared to low AFC animals $(6.9 \pm 5.3 \text{ vs. } 1.9 \pm 2.1; \text{ P}$ < 0.05). Therefore, this study reinforces the evident concept that there are quantitative benefits of embryo production according to the number of antral follicles (Table 1).

Table 1. Reproductive performance of *Bos taurus-indicus* females with high (G-High, follicles \geq 40) or low (L-Low, \leq 10 follicles) antral follicles count comparing the production of embryos *in vitro* (ovum pick up/*in vitro* production - OPU/IVP) and *in vivo* (superovulation (SOV).

Varibles	G-High $(n = 20)$	G-Low $(n = 20)$
Antral follicles (n)	47 ± 6	9 ± 3
Total of oocytes retrieved	738 ^a	116 ^b
Percentage of viable oocytes (%)	58.94 (435/738)	55.17 (64/116)
Cleavage rate (%)	61.25 (452/738)	56.03 (65/116)
Blastocyst rate (%)	16.53 (122/738)	9.48 (11/116)
Total embryos / OPU/IVP (n)	$6.10 \pm 4.51^{aA} (122/20)$	$0.55 \pm 0.83^{bB} (11/20)$
Total structures recovered / collected (n)	$8.80 \pm 6.78^{a} (176/20)$	$2.25 \pm 2.63^{b} (45/20)$
Total embryos / collected (n)	$6.95 \pm 5.34^{aA} (139/20)$	$1.9 \pm 2.13^{\text{bA}} (38/20)$
Frozen proportion (%)	78.42 ^a (109/139)	89.47 ^a (34/38)

The values followed by different superscript letters $(^{a, b})$ within the same line (G-G vs. high-low) or $(^{A, B})$ within the same column (the OPU/IVP vs. SOV) were significantly different (P ≤ 0.05). Adapted from Silva-Santos *et al.* (2014a).

Similar results were obtained with *taurus* donors by Ireland *et al.* (2007), with low AFC females presenting a lower mean number of *in vitro*-produced

embryos compared with high AFC animals (1.3 vs. 4.9 embryos). In *in vivo* production of the embryos, low AFC donors (<15 follicles) produced fewer structures

when compared with high AFC females (>25 follicles), with means of 3.8 vs. 5.4 embryos, respectively.

Contradictory results for indicus animals were found by our research team. In an experiment with Nelore donors (n = 66), they were separated into groups of high AFC (AFC > 40 follicles); medium AFC (18 to 25 follicles) or low AFC (AFC < 7 follicles), and the oocytes were fertilized with sperm from a single bull. In addition to the expected quantitative superiority, a qualitative superiority was observed according to the AFC. Cows with high, medium and low AFC had blastocyst rates of 42, 32 and 13%, respectively (P <0.05; Santos et al., 2015; DCV-CCA-UEL, Londrina, PR, Brazil; unpublished data). However, in a recently concluded study, this pattern was not confirmed. From a larger sample of Nelore cows (n = 356), oocytes were obtained from females of high (>92 follicles), medium (46-76 follicles) or low AFC (<31 follicles). All the oocytes were also fertilized with sperm from a single bull. There was no difference in cleavage and blastocyst rates: 40. 36 and 38% for high, medium and low AFC. respectively (Rosa et al., 2015; DCV-CCA-UEL,

Londrina, PR, Brazil; unpublished data).

In addition to the results of embryo production and pregnancy rates, mRNA expression of genes associated with follicular and oocyte viability seem to show similar patterns in zebu females of high, medium or low AFC. There was no significant difference in the expression of genes related to steroidogenesis (CYP19 and STAR), cell proliferation and differentiation (TGFB1, LIFRa and BMPR2) and hormonal production or response to hormones (AMH, FSHr, PGR and PGRMC) in cumulus and granulosa cells of Nelore females (Rosa *et al.*, 2015; DCV-CCA-UEL, Londrina, PR, Brazil; unpublished data). The contradiction of such results in the same research team reinforces the complexity of this issue.

It is noted that AFC is not the only aspect to quantitatively interfere in embryo production, considering that donors respond to OPU/IVF or SOV according to variables not yet fully understood. In this context, Pontes *et al.* (2009) reported that some donors provided better results on *in vitro* or *in vivo* techniques, regardless of the number of follicles (Table 2).

Table 2. Variations in embryo production rates of six Nelore (*Bos indicus*) donors obtained by *in vitro* procedures (ovum pick up/*in vitro* production - OPU/IVF) or *in vivo* (superovulation and embryo collection - SOV).

Variables	Donor					
variables	Ι	II	III	IV	V	VI
Total procedures OPU/IVF	5	5	4	4	5	5
Mean oocytes per OPU	36.6	25.6	49	29.7	22.8	16
Mean viable oocytes per OPU	32.2	23.4	45.2	26	19.6	14.4
Mean embryos per OPU	15.6	10.4	24.1	10.3	6.8	3.8
Mean pregnancy per OPU	4.8	2.8	9.25	4.3	2.2	1
Total procedures SOV/collection	2	3	2	2	2	3
Mean embryos per SOV	10	4.3	6.5	2	12.5	5.3
Mean pregnancy per SOV	5.5	2	1	1.5	6.5	1.3

Adapted from Pontes et al. (2009).

Despite these individual variations according to the technique, there is a consensus that the quantitative advantages of the high AFC donor should be exploited. Thus, the variation in the number of follicles per donor is currently a very important aspect for the commercial programs of embryos production. A screening method for selection of donors through an ultrasonographic preevaluation is commonly used. The selected donors are generally those with high AFC or a high number of oocytes, which are directly correlated variables. The impact on the final number of pregnancies varies widely, as shown in Table 3.

Table 3. Production of embryos and pregnancies according to the number of oocytes obtained by OPU/IVP (n = 656) from Nelore donors (n = 317). The values (mean \pm SD) are presented per donor.

Donors according to	N°. viable oocytes	N°. viable	N°. pregnancy 30	N°. pregnancy 90
oocyte production		embryos	days	days
Elevated $(n = 78)$	47.06 ± 1.6^{a}	15.06 ± 0.86^{a}	$5.62\pm0.54^{\rm a}$	5.52 ± 0.81^{a}
High $(n = 80)$	24.95 ± 0.33^{b}	9.17 ± 0.63^{b}	3.63 ± 0.36^{b}	3.32 ± 0.33^{b}
Medium $(n=79)$	$15.57 \pm 0.26^{\circ}$	$6.00\pm0.39^{\rm c}$	$2.10 \pm 0.21^{\circ}$	1.92 ± 0.20^{b}
Low (n = 80)	6.31 ± 0.38^{d}	2.42 ± 0.25^{d}	0.92 ± 0.13^{d}	$0.85\pm0.13^{\text{c}}$

^{a-d}Within a column, the mean values with no common superscript differ significantly ($P \le 0.05$). Adapted from Pontes *et al.* (2011).

Despite the unquestionable quantitative advantages in number of embryos for the donor selection method for OPU/IVF based in the AFC, the impact of this criterion on other aspects of fertility and production of meat or milk is not well known for *indicus* animals. Thus, until more studies are performed, the choice of donor based on the number of follicles should be performed only after assessing the genetic merit for production traits of the donor.

Number of antral follicles and anti-Mullerian hormone

AMH is a hormone that belongs to the family of growth factor - β , and is produced by granulosa cells from healthy growing follicles. Its expression is elevated in granulosa cells of small antral follicles and decreases during the follicular growth. The high concentrations of AMH are positively associated with follicular size of mice, women and cows' ovaries population. In the latter, the concentration of AMH is highly correlated with the number of antral follicles and healthy oocytes, thus can be considered a viable endocrine marker of AFC in cattle (Batista *et al.*, 2014).

Bovine females with high AFC (>25 follicles) have higher circulating AMH concentrations in comparison to the females with low AFC (<15 follicles; P < 0.01), with a high correlation between the average concentration of AMH and the mean AFC to Bos taurus cattle (r = 0.88, P < 0.001; Ireland *et al.*, 2008). When comparing taurus and indicus females, both beef and dairy cattle, AMH concentration was also positively correlated with AFC in indicus (Nelore, ranging from 0.56 to 0.68) and taurus (Holstein, ranging from 0.73-0.90). Regardless of the genetic group, females with high AFC showed higher AMH concentrations (0.57 ng/ml in *taurus vs.* 1.20 ng/ml in *indicus*) than females with low AFC (0.06 and 0.78 ng/ml, respectively). Furthermore, there is evidence that AFC in indicus (low and high, 28 and 48 follicles, respectively) is higher than in taurus (low and high, 13 and 34 follicles, respectively; Batista et al., 2014).

Currently, AMH is recognized as a reliable indicator of ovarian response to superovulation. Therefore, the determination of AMH concentration in donor cows may help predict the follicular and ovulatory responses to gonadotropic treatment (Rico *et al.*, 2009).

The AMH concentrations in blood or plasma may be determined by an ELISA test. In a study with Holstein cows over a year, the concentrations of AMH were constant and strongly correlated with the AFC. Donors with AMH concentrations below 87 pg/ml showed less than 15 large follicles by estrous cycle and low efficiency in the production of embryos. Thus, the determination of the concentration of AMH in bovine plasma could routinely be considered in procedures of OPU/IVF and SOV for identification of animals with the best embryo production potential (Rico *et al.*, 2012).

Number of antral follicles and progesterone

The production of progesterone has been linked to physiological activity of the corpus luteum and to ovary and uterus functionality, with direct impact on embryonic development and pregnancy in cattle (Pohler et al., 2012). Low progesterone concentrations are associated with high rates of embryonic mortality, less healthy oocytes and slower growth of the endometrium in these females (Diskin and Morris, 2008). Low AFC taurus females showed low concentrations of progesterone during their oestrus cycle, in comparison with high AFC females. The lower circulating concentrations of progesterone in cows with low AFC were mainly attributed to decreased function of the corpus luteum, possible changes in the responsiveness of luteal cells to LH, a potential reduction STAR protein in the corpus luteum, diminished responsiveness of granulosa and luteal cells to 25-hydroxycholesterol and the reduced ability of granulosa cells of dominant follicles to undergo luteinization in order to produce progesterone (Jimenez-Krassel et al., 2009). It is not clear why an ovulated follicle that turned into a CL would produce less P4 when it is in a low AFC animal. Mainly considering recent studies in Bos indicus in which the situation seems to be the opposite (Seneda et al.. 2015; DCV-CCA-, Londrina, PR, Brazil; unpublished data). Considering this and other aspects, we believe in distinct pathways in Bos taurus and Bos indicus at least for some reproductive aspects. It has also been demonstrated that high AFC was positively associated with endometrial thickness. An increased endometrial thickness was associated with higher embryonic implantation rates (Basir et al., 2002).

Antral follicle count and fertility parameters in beef heifers and cows

The low AFC in dairy *taurus* females (Holstein) was associated with several characteristics related to unfertility, such as smaller ovaries (Ireland *et al.*, 2008), less chance of pregnancy at the end of the breeding season (Mossa *et al.*, 2012), reduced responsiveness to the SOV treatment, fewer viable embryos (Singh *et al.*, 2004; Ireland *et al.*, 2007), lower circulating concentrations of progesterone and AMH (Ireland *et al.*, 2011; Evans *et al.*, 2012) and reduced endometrial thickness (Jimenez-Krassel *et al.*, 2009). In this context, animals with high AFC were proved superior in all of the mentioned aspects. Thus, it is assumed and substantiated that in Holstein cows a linear correlation exists between high AFC and indicators for reproductive efficiency.

This context, however, has not been proven in Zebu animals. Recent studies have shown that reproductive traits of *indicus* (Nelore) and *taurus-indicus* females (Braford) did not show improved performance associated with high AFC (Santos *et al.*, 2012, 2013; Mendonça *et al.*, 2013; Morotti *et al.*, 2014;

Santos *et al.*, 2015, DCV-CCA-UEL, Londrina, PR, Brazil, unpublished data). A higher rate of follicular growth and larger follicular diameters have been described in *indicus-taurus* (Santos *et al*, 2012) and *indicus* (Morotti *et al*, 2014) females with low-AFC. Also, dominant follicles with larger diameters have been described in Nelore females with low AFC (Morotti *et al.*, 2015; DCV-CCA-UEL, Londrina, PR, Brazil; unpublished data).

Considering the AFC bimonthly basis in 137 *taurus-indicus* females, there was no difference in the average body weight from nine to 24 months, or in the diameter of the uterine horn (Santos *et al.*, 2012). In a subsequent experiment, 71 females from same herd were subjected to a TAI protocol to determine the values of certain variables associated with follicular dynamics. The data are presented in Table 4.

Table 4. Mean \pm SD values of follicular dynamics of *taurus-indicus* females (Braford) with different antral follicles counts (AFC; high, \geq 40 follicles; medium, 17-23 follicles; and low, \leq 10 follicles) after a protocol of ovulation synchronization.

Variables	High ≥ 40 follicles n = 24	Medium 17-23 follicles n = 24	$Low \le 10 \text{ follicles} \\ n = 23$
Antral follicle count on D5 (n)	47 ± 9.9^{a}	24 ± 9.9^{b}	$9\pm3.9^{\circ}$
Diameter of the largest follicle on D5 (cm)	0.66 ± 0.3	0.70 ± 0.2	0.80 ± 0.2
Ovulatory follicle diameter (cm)	1.15 ± 0.2^{a}	1.27 ± 0.2^{b}	1.32 ± 0.2^{b}
Interval between device removal and ovulation (h)	69.33 ± 5.1	71.25 ± 3.0	70.50 ± 4.1
Ovulation rate (%)	75 (18/24)	67 (16/24)	70 (16/23)
CL diameter (cm)	1.93 ± 0.3	1.97 ± 0.3	2.04 ± 0.3

^{a-c}Within a column, the mean values with no common superscript differ significantly ($P \le 0.05$). Adapted from Santos *et al.* (2012).

Unlike the results reported for Holstein cows (Ireland *et al.*, 2011; Evans *et al.*, 2012), the data in Table 4 show that Braford females of low and medium AFC had ovulatory follicles with greater diameters.

Regarding pregnancy rates of *indicus* (Nelore) females, two studies have shown pregnancy data after TAI, one with postpartum cows (n = 691) and the other with heifers at 24 months of age (n = 208). On day 8 of the TAI protocol, the ovaries were evaluated by

ultrasonography and females were classified into groups of high, medium or low AFC. Pregnancy rates were evaluated by transrectal ultrasonography 30 days after TAI. The data, summarized in Table 5, demonstrate that low AFC cows were superior to medium AFC cows, while high AFC cows were similar to both. There were no differences for heifers. In short, it was not possible to establish the same relationship described for *taurus* females.

Table 5. Reproductive performance of Nelore (*Bos indicus*) females with high, medium and low antral follicle count (AFC) after a timed artificial insemination protocol.

Cows (n)	High ≥25 follicles (149)	Medium 11-24 follicles (400)	Low ≤10 follicles (142)	Total (691)
Antral follicles (n)	30.70 ± 5.66^{a}	17.03 ± 3.28^{b}	$7.83 \pm 2.42^{\circ}$	17.93 ± 8.45^{b}
Pregnancy rate (%)	51.67 ^{ab}	48.00 ^b	60.50^{a}	51.49 ^{ab}
Heifers (n)	High ≥30 follicles (38)	Medium 13-29 follicles (143)	Low ≤12 follicles (27)	Total (208)
Antral follicles (n)	37.73 ± 7.05^a	19.23 ± 4.29^{b}	$10.55 \pm 2.17^{\circ}$	21.48 ± 9.47^{b}
Pregnancy rate (%)	44.73	43.35	51.85	44.71

^{a-c}Within a column, the mean values with no common superscript differ significantly ($P \le 0.05$). Data are presented as mean ± SD. Adapted from Santos *et al.* (2013) and Mendonça *et al.* (2013).

The association between AFC and pregnancy rates was also analyzed in *indicus* recipients after the transfer of *in vitro*-produced embryos. Cyclical Nelore heifers (n = 281, ECC 3.0 ± 0.5) underwent a classic

protocol for ovulation synchronization before TET. The heifers received the embryos 17 days after the beginning of the treatment. There was no difference in pregnancy rate according to AFC (Table 6).

High	Medium	Low
\geq 25 follicles	11 follicles	\leq 3 follicles
(38)	(136)	(75)
$25.8 \pm 7,4^{a}$	11.3 ± 2.9^{b}	3.8 ± 1.3^{c}
30.0	33.8	34.6
	$\frac{\geq 25 \text{ follicles}}{(38)}$ $25.8 \pm 7,4^{a}$	

Table 6. Antral follicle count and pregnancy rates of Nelore (*Bos indicus*) recipients after TET of *in vitro* embryo production.

^{a-c}Within a column, the mean values with no common superscript differ significantly ($P \le 0.05$).

Antral follicle count, carcass phenotype and genetic characteristics of heritability

very hard to compare data.

In a recent study, heritability and the impact of environmental effects during pregnancy on AFC was evaluated in cattle. In the Holstein breed, this parameter had heritability of 0.31 ± 0.14 and 0.25 ± 0.13 for cows and heifers, respectively. The AFC was negatively associated with genetic merit for milk fat. The authors concluded that the AFC in Holstein females is an inherited genetic trait moderately affected by age and lactation status, but not the mother's milk production level during pregnancy (Walsh *et al.*, 2014).

To *indicus-taurus* heifers (Braford, n = 270), the AFC was considered in a statistical model associated with phenotypic characteristics for selecting matrices. The variables considered were mean number of antral follicles, effect of contemporary group, age, birth weight gain at weaning, conformation at weaning, finishing at weaning precocity, musculature at weaning, weight gain from weaning to yearling, forming at yearling, precocity at yearling and musculature at yearling. For all parameters studied, the correlation values were very low (0.056 to 0.082; P > 0.05), pointing out that, for taurus-indicus animals, the antral follicle population has no association with the main selection criteria of matrices for beef herds (Morotti et al., 2015; DCV-CCA-UEL, Londrina, PR, Brazil; unpublished data).

Final comments

After all comments above, our general conclusion is: so far, it is not possible to apply the *Bos taurus* AFC model for *Bos indicus* females. While the AFC seems to be a very clear criterium for reproductive selection in *taurus*, we have found contradictory results in *indicus* cattle. There are results signaling for a better reproductive efficiency of low AFC animals, others to the medium AFC females and even, in the same model of *taurus* cows, data suggesting that there is a greater reproductive efficiency in high AFC zebu females.

One of the main challenges in the analysis of articles refers to the large variation of the classification criteria of high, medium or low follicle count. There are values considered as low AFC in one article that are considered as high AFC in other study, using the same category and the same breed. This situation makes it Nevertheless, the inconstancy of the results shows the need to always prioritize genetic merit in the choice of a donor, and not AFC. Following this criterion is particularly important for oocyte donors. In *in vitro* embryo production, the quantitative advantage of number of structures tends to create a biased selection of donors with high AFC, because they make the entire IVEP chain more profitable. Considering the estimate that Brazil, as the world leader of this biotechnology, produces annually around 300,000 embryos *in vitro* (Viana *et al.*, 2012) it is necessary to consider that an immediate benefit in the number of embryos should not supercede a strict criterion of genetic merit, which is a more important aspect in short, medium and long term in the selection of animals.

References

Basir GS, O W-S, So WWK, Ng EHY, Ho PC. 2002. Evaluation of cycle-tocycle variation of endometrial responsiveness using transvaginalsonography in women undergoing assisted reproduction. *Ultrasound Obstet Gynecol*, 19:484-489.

Batista EOS, Macedo GG, Sala RV, Ortolan MDDV, Sá Filho MF, Del Valle TA, Jesus EF, Lopes RNVR, Renn FP, Baruselli PS. 2014. Plasma Antimullerian hormone as a predictor of ovarian antral follicular population in *Bos indicus* (Nelore) and *Bos taurus* (Holstein) heifers. *Reprod Domest Anim*, 49:448-452.

Boni R. 2012. Ovum pick-up in cattle: a 25 yr retrospective analysis. *Anim Reprod*, 9:362-369.

Burns DS, Jimenez-Krassel F, Ireland JLH, Knight PG, Ireland JJ. 2005. Numbers of antral follicles during follicular waves in cattle: evidence for high variation among animals, very high repeatability in individuals, and an inverse association with serum follicle-stimulating hormone concentrations. *Biol Reprod*, 73:54-62.

Cushman RA, Allan MF, Kuehn LA, Snelling WM, Cupp AS, Freetly HC. 2009. Evaluation of antral follicle count and ovarian morphology in crossbred beef cows: investigation of influence of stage of the estrous cycle, age, and birth weight. *J Anim Sci*, 87:1971-1980.

Diskin MG, Morris DG. 2008. Embryonic and early foetal losses in cattle and other ruminants. *Reprod Domest Anim*, 43:260-267.

Evans ACO, Mossa F, Walsh SW, Scheetz D, Jimenez-

Krassel F, Ireland JLH, Smith GW, Ireland JJ. 2012. Effects of maternal environment during gestation on ovarian folliculogenesis and consequences for fertility in bovine offspring. *Reprod Domest Anim*, 47:31-37.

Hansen P. 2014. Current and Future Assisted Reproductive Technologies for Mammalian Farm Animals. *In*: Lamb GC, DiLorenzo N (Ed.). *Current and Future Reproductive Technologies and World Food Production*. New York: Springer. pp. 1-22.

Hehenkamp WJK, Looman CWN, Themmen APN, De Jong FH, Te Velde ER, Broekmans FJM. 2006. Anti-Mullerian hormone levels in the spontaneous menstrual cycle do not show substantial fluctuation. *J Clin Endocrinol Metab*, 91:4057-4063.

Ireland JJ, Ward F, Jimenez-Krassel F, Ireland JLH, Smith GW, Lonergan P, Evans ACO. 2007. Follicle numbers are highly repeatable within individual animals but are inversely correlated with FSH concentrations and the proportion of good-quality embryos after ovarian stimulation in cattle. *Hum Reprod*, 22:1687-1695.

Ireland JLH, Scheetz D, Jimenez-Krasse F, Themmen APN, Ward F, Lonergan P, Smith GW, Perez GI, Evans ACO, Ireland JJ. 2008. Antral follicle count reliably predicts number of morphologically healthy oocytes and follicles in ovaries of young adult cattle. *Biol Reprod*, 79:1219-1225.

Ireland JJ, Zielak AE, Jimenez-Krassel F, Folger J, Bettegowda A, Scheetz D, Walsh S, Mossa F, Knight PG, Smith GW, Lonergan P, Evans ACO. 2009. Variation in the ovarian reserve is linked to alterations in intrafollicular oestradiol production and ovarian biomarkers of follicular differentiation and oocyte quality in cattle. *Biol Reprod*, 80:954-964.

Ireland JJ, Smith GW, Scheetz D, Jimenez-Krassel F, Folger JK, Ireland JLH, Mossa F, Lonergan P, Evans ACO. 2011. Does size matter in females? An overview of the impact of the high variation in the ovarian reserve on ovarian function and fertility, utility of anti-Müllerian hormone as a diagnostic marker for fertility and causes of variation in the ovarian reserve in cattle. *Reprod Fertil Dev*, 23:1-14.

Jimenez-Krassel F, Folger JK, Ireland JL, Smith GW, Hou X, Davis JS, Lonergan P, Evans AC, Ireland JJ. 2009. Evidence that high variation in ovarian reserves of healthy Young adults has a negative impact on the corpus luteum and endometrium during estrous cycles in cattle. *Biol Reprod*, 80:1272-1281.

Mapletoft RJ, Hasler JF. 2005. Assisted reproductive technologies in cattle: a review. *Rev Sci Tech*, 247:393-403.

Mendonça HGR, Santos GMG, Silva-Santos KC, Barreiros TRR, Blaschi W, Morotti F, Moraes FLZ, Seneda MM. 2013. Pregnancy rates following FTAI of Nelore heifers (*Bos indicus*) with high, intermediate and low numbers of antral follicles. *Anim Reprod*, 10:452. (abstract). Morotti F, Santos GMG, Silva-Santos KC, Moritz Y, Marcantonio TN, Seneda MM. 2014. Follicular dynamics in Nelore females (*Bos indicus*) with high or low numbers of antral follicles. *Anim Reprod*, 11:352. (abstract).

Mossa F, Jimenez-Krassel F, Folger JK, Ireland JLH, Smith GW, Lonergan P, Evans AC, Ireland JJ. 2010. Evidence that high variation in antral follicle count during follicular waves is linked to alterations in ovarian androgen production in cattle. *Reproduction*, 140:713-720.

Mossa F, Walsh SW, Butler ST, Berry DP, Carter F, Lonergan P, Smith GW, Ireland JJ, Evans ACO. 2012. Low numbers of ovarian follicles ≥ 3 mm in diameter are associated with low fertility in dairy cows. *J Dairy Sci*, 95:2355-2361.

Pohler KG, Geary TW, Atkins JA, Perry GA, Jinks EM, Smith MF. 2012. Follicular determinants of pregnancy establishment and maintenance. *Cell Tissue Res*, 349:649-664.

Pontes JHF, Nonato-Junior I, Sanches BV, Ereno-Junior JC, Uvo S, Barreiros TRR, Oliveira JA, Hasler JF, Seneda MM. 2009. Comparison of embryo yield and pregnancy rate between in vivo and in vitro methods in the same Nelore (*Bos indicus*) donor cows. *Theriogenology*, 71:690-697.

Pontes JHF, Silva KCF, Basso AC, Rigo AG, Ferreira CR, Santos GMG, Sanches BV, Porcionato JPF, Vieira PHS, Faifer FS, Sterza FAM, Schenk JL, Seneda MM. 2010: Large-scale in vitro embryos production and pregnancy rates from *Bos taurus*, *Bos indicus*, and *indicus-taurus* dairy cows using sexed sperm. *Theriogenology*, 74:1349-1355.

Pontes JHF, Melo-Sterza FA, Basso AC, Ferreira CR, Sanches BV, Rubin KCP, Seneda MM. 2011. Ovum pick up, in vitro embryo production, and pregnancy rates from a large-scale commercial program using Nelore cattle (*Bos indicus*) donors. *Theriogenology*, 75:1640-1646.

Rico C, Fabre S, Médigue C, Di Clemente N, Clément F, Bontoux M, Touzé JL, Dupont M, Briant E, Rémy B, Beckers JF, Monniaux D. 2009. Antimullerian hormone is an endocrine marker of ovarian gonadotropin responsive follicles and can help to predict superovulatory responses in the cow. *Biol Reprod*, 80:50-59.

Rico C, Drouilhet L, Salvetti P, Dalbiès-Tran R, Jarrier P, Touzé J-L, Pillet E, Ponsart C, Fabre S, Monniaux D. 2012. Determination of anti-Müllerian hormone concentrations in blood as a tool to select Holstein donor cows for embryo production: from the laboratory to the farm. *Reprod Fertil Dev*, 24:932-944.

Santos GMG, Silva-Santos KC, Siloto LS, Morotti F, Marcantonio TN, Marinho LSR, Thasmo RLO, Koetz JRC, Cintra DML, Seneda MM. 2012. Dinâmica folicular em fêmeas bovinas de alta, média e baixa contagem de folículos antrais: resultados preliminares. *Acta Sci Vet*, 40:422. (abstract). Santos GMG, Silva-Santos KC, Barreiros TRR, Blaschi W, Morotti F, Silva CB, Moraes FLZ, Giudicissi DPL, Seneda MM. 2013. Conception rates following FTAI of Nelore cows (*Bos indicus*) with high, intermediate and low numbers of antral follicles. *Anim Reprod*, 10:451. (abstract).

Santos GMG, Silva-Santos KC, Barreiros TRR, Morotti F, Sanches, BV, Moraes FLZ, Blaschi W, Seneda MM. 2014. High numbers of antral follicles influence the in vitro embryo production, but not the conception rate of fixed-time artificial insemination in nelore cattle. *Reprod Fertil Dev*, 27:206. (abstract).

Silva-Santos KC, Santos GMG, Siloto LS, Hertel MF, Andrade ER, Rubin MIB, Sturion L, Melo-Sterza FA, Seneda MM. 2011. Estimate of the population of preantral follicles in the ovaries of *Bos taurus indicus* and *Bos taurus taurus* cattle. *Theriogenology*, 76:1051-1057.

Silva-Santos KC, Santos GMG, Koetz Júnior C, Morotti F, Siloto LS, Marcantonio TN, Urbano MR, Oliveira RL, Lima DCM, Seneda MM. 2014a. Antral follicle populations and embryo production - in vitro and in vivo - of *Bos indicus-taurus* donors from weaning to yearling ages. *Reprod Domest Anim*, 49:228-232. Silva-Santos KC, Siloto LS, Santos GMC, Morotti F, Marcantonio TN, Seneda MM. 2014b. Comparison of antral and pre-antral ovarian follicle populations between *Bos indicus* and *Bos indicus-taurus* cows with high or low antral follicles counts. *Reprod Domest Anim*, 49:48-51.

Singh J, Dominguez M, Jaiswal R, Adams GP. 2004. A simple ultrasound test to predict the superstimulatory response in cattle. *Theriogenology*, 62:227-243.

Viana JHM, Siqueira LGB, Palhao MP, Camargo LSA. 2012. Features and perspectives of the Brazilian in vitro embryo industry. *Anim Reprod*, 9:12-18.

Taneja M, Bols PE, Van De Velde A, Ju JC, Schreiber D, Tripp MW, Levine H, Echelard Y, Riesen J, Yang X. 2000. Developmental competence of juvenile calf oocytes in vitro and in vivo: influence of donor animal variation and repeated gonadotrophin stimulation. *Biol Reprod*, 62:206-213.

Walsh SW, Mossa F, Butler ST, Berry DP, Scheetz D, Jimenez-Krassel,F, Tempelman RJ, Carter F, Lonergan P, Evans ACO, Ireland JJ. 2014. Heritability and impact of environmental effects during pregnancy on antral follicle count in cattle. *J Dairy Sci*, 97:4503-4511.